

LEAD ACETATE AND LEAD NITRATE INDUCED SEX-LINKED RECESSIVE LETHAL MUTATIONS IN MATURE SPERM OF *Drosophila melanogaster*

ROSARIO RODRIGUEZ-ARNAIZ, PATRICIA
RAMOS MORALES AND MARÍA ISABEL
CONCHESO COBOS,

Laboratorio de Genética, Facultad de Ciencias,
Universidad Nacional Autónoma de México, Co-
yoacán 04510, México, D. F.

ABSTRACT

The mutagenic effects of two lead salts were investigated: acetate and nitrate on mature sperm of *Drosophila melanogaster*. Oregon R adult males were injected intraperitoneally with different concentrations of the salts: 650, 750 and 850 ppm for lead acetate and 25, 50 and 75 ppm for lead nitrate. Treated males were mated with virgin BASC females and the F_2 scored for the induction of sex-linked recessive lethal mutations. The higher doses of both salts were found to induce a significant number of these mutations at 5% level on the Kastenbaum-Bowman test (1970), lead acetate being less toxic and less mutagenic than lead nitrate.

RESUMEN

Se investigaron los efectos mutagénicos de dos sales de plomo: acetato y nitrato en espermatozoides maduros de *Drosophila melanogaster*. Machos adultos Oregon R fueron inyectados intraperitonealmente con diferentes concentraciones de las sales: 650, 750 y 850 ppm de acetato de plomo y 25, 50 y 75 ppm de nitrato de plomo. Los machos tratados fueron cruzados con hembras vírgenes BASC y la F_2 fue examinada para valorar la inducción de mutaciones letales recesivas ligadas al sexo. Ambas sales produjeron estas mutaciones en un nivel de 5% en la prueba de Kastenbaum-Bowman (1970), siendo el acetato de plomo menos tóxico y menos mutagénico que el nitrato de plomo.

INTRODUCTION

Drosophila melanogaster has been used for mutagenic studies ever since H. J. Müller developed the sex-linked recessive lethal test (SLRL) to detect the mutagenic effects of X rays (Muller 1927). He used the C1B chromosome which was later replaced by the Muller-5 or BASC chromosome. Using *Drosophila*, Auerbach and Robson (1946) were the first to discover the mutagenic effects of mustard gas. Since then a great number of chemicals have been tested for mutagenic activity in *Drosophila* (Lee *et al.* 1983).

Among the heavy metals, lead, has been shown to be widely distributed in the atmosphere, water, soils and foods (Beliles 1975). At the cellular level, lead inhibits the activity of enzymes that are dependant on the presence of free sulphhydryl groups (SH). The clearest manifestation of these effects is the disturbance on the biosynthesis of heme, which in humans is accompanied by abnormalities in porphyrin metabolism (Valle and Ulmer 1972).

Lead acetate is used as a topical astringent and is found to be a renal carcinogen in rats (Bolyland *et al.* 1962, Van Esch *et al.* 1962, Roe *et al.* 1965, Mao and Molnar 1967, Choie and Richter 1974, Furst *et al.* 1976). In the Syrian hamster lead induces neoplastic changes in the bronchio-alveolar area (Kobayashi and Okamoto 1974, ICPEMC 1984). It also produces infertility in mice (Varma *et al.* 1974) and reduces the reproductive ability of rats (Stowe and Goyer 1971, Hackett *et al.* 1982, Hess and Sikov 1982).

Lead nitrate is employed in the manufacture of matches and explosives, as a mordant in dyeing of textiles, as well as a sensitizer in photography. It is embryotoxic to rats (McClain and Becker 1975) but not teratogenic (McClain and Siekier 1975). In *Drosophila melanogaster* lead induces enzymatic alterations in esterase and triose phosphate isomerase (Lower *et al.* 1976) and affects non disjunction (Ramel 1973).

Information about the mutagenic effects of lead salts in humans who are occupationally exposed to them and information obtained from in vitro studies are contradictory (Maki-Paakkanen *et al.* 1981).

We report the mutagenic effects of two lead salts, namely, acetate and nitrate on the induction of SLRL in mature sperm of *Drosophila melanogaster*.

MATERIALS AND METHODS

Lead acetate $[(CH_3COO)_2Pb \cdot 3H_2O]$ (Baker) was employed at concentrations of 650, 750 and 850 ppm. Lead nitrate $[Pb(NO_3)_2]$ (Baker) was used at concentrations of 25, 50 and 75 ppm. The highest concentration of both salts is their LD_{50} . The salts were dissolved in distilled water.

Oregon R adult males were injected intraperitoneally (Felix and Rodriguez-Arnaiz 1968) with almost $0.2 \mu l$ of the salts solutions (Sega and Lee 1970) and after treatment the males were mated with BASC virgin females in a proportion of one male to two females, for three days. F_1 females fertilized by F_1 males were put into fresh medium, one per vial, and the F_2 scored for the induction of SLRL mutations by means of the absence of wild males. Only mature sperm were sampled (Vogel 1975). Controls were injected with $0.2 \mu l$ of 5% sucrose solution (Vogel and Sobels 1976). The Kastenbaum-Bowman test (1970) was employed in order to calculate the statistical significance of the results.

RESULTS AND DISCUSSION

Chemical agents which induce genetic damage constitute a risk for all populations, including human beings. The sex-linked recessive lethal test is capable of detecting chemicals which are capable of inducing heritable lethal mutations which may range from changes in only one base of the DNA, that is a point mutation, to small

deletions or chromosome aberrations. The SLRL test simultaneously examines 600 to 800 genes, which represents about 80% of the loci of the X-chromosome or roughly one fifth of the *Drosophila* genome (Lee *et al.* 1983).

Both lead salts induced SLRL (Table I), lead acetate was less toxic and less mutagenic than lead nitrate.

TABLE I. INDUCTION OF SEX-LINKED RECESSIVE LETHALS
BY LEAD ACETATE AND LEAD NITRATE ON MATURE
SPERM OF *Drosophila melanogaster*

Concentration (ppm)	Number of tested chromosomes	Number of induced lethals	Percentage of lethals
Control	1 178	7	0.59
Lead acetate			
650	1 285	11	0.86
750	1 061	10	0.94
850	1 327	28	2.11 *
Lead nitrate			
25	1 205	15	1.24
50	1 326	21	1.58 *
75	1 584	34	2.15 *

* Significant at 5% level on Kastenbaum-Bowman test (1970).

In the *Tradescantia* micronucleus test, the mutagenic effect of lead nitrate was found to be significant while lead acetate gave a borderline response (Ma *et al.* 1984).

In cattle, lead does not produce severe chromosome abnormalities (Leonard *et al.* 1974). In monkeys with a low calcium diet, lead acetate induces dicentrics, translocations and exchanges, while in groups receiving lead, irrespective of the diet, lead acetate produce gaps and fragments (Deknudt *et al.* 1977). In humans, reports about the induction of chromosomal aberrations in lead exposed individuals are contradictory. Some have reported positive results (Garza-Chapa *et al.* 1977, Hogshead *et al.* 1979, Forni *et al.* 1980) while others have not found any increase in rates of chromosome aberrations (Schmid and Bauchinger 1973, O'Riordan and Evans 1974, Bijlsma and De France 1976, Horvat and Prpić-Majić 1977, Maki-Paakkanen *et al.* 1981. Qazi *et al.* (1980) observed a temporary increase in chromosome breakage in an infant exposed to lead while in utero. In some experiments, human lymphocytes exposed to lead salts in vitro, have been reported as unaffected (Deknudt and Deminatti 1978, Bauchinger and Gasiorek 1980), although Beek and Oboe (1974) obtained increased frequencies of chromosome aberrations.

In *Drosophila* we have found a statistical significant increase in the frequency of SLRL mutations. However other experiments are needed to show the mode of action of these mutagens (Valencia *et al.* 1984).

REFERENCES

- Auerbach C. and Robson J. M. (1946). Chemical production of mutations. *Nature* (London) 157, 302.
- Bauchinger M. and Gasiorek K. (1980). Chromosome analysis on human lymphocytes after combined treatment with lead, cadmium and zinc. 10th Annual Meeting of EMMS, Greece. Abstracts p. 55.
- Beek B. and Obe G. (1974). Effects of lead acetate on human leucocyte chromosomes in vitro. *Experientia* 30, 1006-1007.
- Beliles R. P. (1975). Metals. In *Toxicology*. Casarett, L. J. and Doull J. Eds. Macmillan Pub. New York pp. 477-482.
- Bijlsma J. B. and De France H. F. (1976). Cytogenetic investigations in volunteers ingesting inorganic lead. *Int. Arch. Occup. Environ. Health* 38, 145-148.
- Boyland E., Dukes C. E., Grover P. L. and Mitchley B. C. V. (1962). The induction of renal tumours by feeding lead acetate to rats. *Brit. J. Cancer* 16, 283-288.
- Choie D. D. and Richter G. W. (1974). Cell proliferation in mouse kidney induced by lead. 1 Synthesis of DNA. *Lab. Invest.* 30, 647-651.
- Deknadt G. H., Colle A. and Gerber G. B. (1977). Chromosomal abnormalities in lymphocytes from monkeys poisoned with lead. *Mutat. Res.* 45, 77-83.
- and Deminatti M. (1978). Chromosome studies in human lymphocytes after in vitro exposure to metal salts. *Toxicology* 10, 67-75.
- Felix E. R. and Rodriguez-Arnaiz R. (1968). A microinjection technique for *Drosophila*. *D. Inform. Service* 43, 180.
- Forni A., Sciamé A., Bertazzi P. A. and Alessio L. (1980). Chromosome and biochemical studies in women occupationally exposed to lead. *Arch. Environ. Health* 35, 139-145.
- Furst A., Schlauder M. and Sasmore D. P. (1976). Tumorigenic activity of lead chromate. *Cancer Res.* 36, 1779-1783.
- Garza-Chapa R., Leal-Garza C. H. and Molina-Ballesteros G. (1977). Análisis cromosómico en personas profesionalmente expuestas a contaminación por plomo. *Arch. Invest. Med.* 8, 11-20.
- Hackett P. L., Hess J. O. and Sikov M. R. (1983). Effect of dose level and pregnancy on the distribution and toxicity of intravenous lead in rats. *J. Tox. Environ. Health* 9, 1007-1020.
- Hess J. O. and Sikov M. R. (1982). Distribution and effects of intravenous lead in the fetoplacental unit of the rat. *J. Tox. Environ. Health* 9, 1021-1032.
- Hogstedt B., Kolnif A. M., Mitelman I. and Shutz A. (1979). Correlation between blood-lead and chromosomal aberrations. *Lancet* (2) 8136.
- Horvat D. J. and Prpé-Majié D. (1977). Cytogenetic and biochemical study of lead exposed populations, 2nd International Conference on Environmental Mutagens, Edinburgh, England Abstracts p. 174.
- ICPEMC (1984). Report of ICPEMC task group 5 on the differentiation between genotoxic and non-genotoxic carcinogens. *Mutat. Res.* 133, 1-49.
- Kastenbaum M. A. and Bowman K. O. (1970). Tables for determining the statistical significance of mutation frequencies. *Mutat. Res.* 9, 527-549.
- Kobayashi N. and Okamoto T. (1974). Effects of lead oxides on the induction of lung tumours in Syrian Hamster. *J. Natl. Cancer Inst.* 52, 1605-1607.
- Lee W. R., Abrahamson S., Valencia R., Von Halle E. S., Wurgler F. E. and Zimmering S. (1983). The sex-linked recessive lethal test for mutagenesis in *Drosophila melanogaster*. A report of the U. S. Environmental Protection Agency Gene-Tox Program. *Mutat. Res.* 123, 183-279.
- Léonard A., Deknadt G. H. and Debackere M. (1974). Cytogenetic investigations on leucocyte of cattle intoxicated with heavy metals. *Toxicology* 2, 269-273.
- Lower W. F., Drobney V. K., Rose P. S. and Putnam C. W. (1976). Environmental and laboratory monitoring of biotic indicators of heavy metals. *Mutat. Res.* 38, 386.
- Maki-Paakkanen J. M., Sorsa M. and Vainio H. (1981). Chromosome aberrations and sister chromatid exchanges in lead exposed workers. *Hereditas* 94, 269-275.
- Ma T. H., Harris M. M., Anderson V. A., Ahmed I., Mohammad K., Bare J. L., and Lin G. (1984). *Tradescantia* micronucleus (Trad-MCN) test on 140 health-related agents. *Mutat. Res.* 138, 157-167.

- Mao P. and Molnar J. J. (1967). The fine structure and histochemistry of lead induced renal tumors in rats. *Am. J. Pathol.* 50, 571-580.
- McClain R. M. and Becker B. A. (1975). Teratogenicity, fetal toxicity and placental transfer of lead nitrate in rats. *Toxicol. Appl. Pharmacol.* 31, 72-82.
- and Sierkierka J. J. (1975). The effects of various chelating agents on the teratogenicity of lead nitrate in rats. *Toxicol. Appl. Pharmacol.* 31, 434-442.
- Muller H. J. (1927). Artificial transmutation of the gene. *Science* 66, 84-87.
- O'Riordan M. L. and Evans H. J. (1974). Absence of significant chromosome damage in males occupationally exposed to lead. *Nature (London)* 247, 50-53.
- Qazi Q. H., Madahar C. and Yuceoglu A. M. (1980). Temporary increase in Chromosome breakage in an infant prenatally exposed to lead. *Human. Gen.* 53, 201-203.
- Ramel C. (1973). Effects of metal compounds on chromosome segregation. *Mutat. Res.* 21, 45-46.
- Roe F. J. C., Boyland E., Dukes C. E. and Mitchley B. C. V. (1965). Failure of testosterone or xanthopterin to influence the induction of renal neoplasms by lead in rats. *Brit. J. Cancer* 19, 860-866.
- Schmid E. and Bauchinger M. (1973). Chromosome analysis in mammalian cells after exposure to lead in vitro and in vivo. *Mutat. Res.* 21, 48.
- Sega G. A. and Lee R. W. (1970). A vacuum injection technique for obtaining uniform dosage in *Drosophila melanogaster*. *D. Inform. Service* 45, 179.
- Stowe H. D. and Goyer R. A. (1971). The reproductive ability and progeny of F₁ lead toxic rats. *Fertility and Sterility* 22, 755-760.
- Valencia R., Abrahamson S., Lee W. R., Von Halle E. S., Woodruff R. C., Wurgler F. E. and Zimmering S. (1984). Chromosome mutation test for mutagenesis in *Drosophila melanogaster*. A report of the U. S. Environmental Protection Agency Gene-Tox Program. *Mutat. Res.* 134, 61-88.
- Valle B. L. and Ulmer D. D. (1972). Biochemical effects of mercury, cadmium and lead. *Ann. Rev. Biochem.* 41, 91-128.
- Van Esch G. J., Gendersen H. and Vink H. H. (1962). The induction of renal tumors by feeding of basic lead acetate to rats. *Brit. J. Cancer* 16, 289-297.
- Varma M. M., Joshi S. R. and Adeyami A. O. (1974). Mutagenicity and infertility following administration to lead sub-acetate to swiss male mice. *Experientia* 30, 486-487.
- Vogel E. (1975). Some aspects of the detection of potential mutagen agents in *Drosophila*. *Mutat. Res.* 29, 241-250.
- and Sobels F. H. (1976). The function of *Drosophila* in genetic toxicology testing. In *Chemical mutagens: principles and methods for their detection*. (A. Hollaender, Ed.). Plenum Press, New York, Vol. 4, pp. 93-142.